

1 ***Streptomyces xinghaiensis* sp. nov., a new actinomycete derived from marine**  
2 **sediment sample in Xinghai Bay, Dalian, China**

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21 **Running title:** *Streptomyces xinghaiensis* sp.nov.

22 **Category:** New Taxa - Actinobacteria

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24 The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain  
25 S187<sup>T</sup> is **EF577247**.

26 A novel actinomycete strain S187<sup>T</sup> was isolated from a marine sediment sample collected  
27 from Xinghai Bay, Dalian, China. Growth occurred on ISP medium 2 with 0-9% NaCl,  
28 pH 6.0-9.0 and 10-45 °C. The cell wall of strain S187 contained **LL**-Diaminopimelic acid  
29 (DAP) isomer. The predominant menaquinones were MK-9 (H<sub>6</sub>) (40.8%), MK-9 (H<sub>8</sub>)  
30 (38.2%), and MK-9 (H<sub>2</sub>) (8.8%). The major fatty acids were **iso-C<sub>16:0</sub> (29.6%),**  
31 **anteiso-C<sub>15:0</sub> (14.0%) and anteiso-C<sub>17:0</sub> (11.6%).** Phospholipids of the cells contained  
32 phosphatidylethanolamine, phosphatidylinositol, phosphatidylglycerol,  
33 phosphatidylinositol mannosides and one unknown phospholipid. The G+C content of  
34 the genomic DNA was 72.01 mol%. 16S rRNA gene sequence of the isolate had 98.1%  
35 sequence similarity with *Streptomyces flavofuscus* NRRL B-8036 (=DSM 41426<sup>T</sup>) and  
36 97.5% similarity with *Streptomyces albiaxis* DSM 41799<sup>T</sup>, respectively, showing that  
37 the new isolate should be assigned to the genus *Streptomyces*. Meanwhile, DNA–DNA  
38 hybridizations against the above-mentioned two *Streptomyces* species showed 31.4% and  
39 46.9% relatedness, respectively. Moreover, the three strains differed in some  
40 physiological and biochemical properties. Thus, on the basis of phenotypic and genotypic  
41 analyses, it is proposed that strain S187<sup>T</sup> represents a novel species of the genus  
42 *Streptomyces*, for which the name *Streptomyces xinghaiensis* sp. nov. is proposed with the  
43 type strain S187<sup>T</sup> (=NRRL B24674<sup>T</sup>= CCTCC AA 208049<sup>T</sup>= KCTC 19546<sup>T</sup>).

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49 Marine actinobacteria have been screened and identified intensively in recent years  
50 (Maldonado *et al.*, 2005; Ward & Bora, 2006). It was demonstrated that a diverse assemblage  
51 of actinobacteria are present in marine environment, including marine sediment (Jensen *et al.*,  
52 2005; Bredholdt *et al.*, 2007), especially deep sea sediment (Pathom-Aree *et al.*, 2006), as  
53 well as marine sponges (Montalvo *et al.*, 2005; Zhang *et al.*, 2006). Dalian is a costal city  
54 located in the southernmost of Liaodong Peninsula of China, and has a very long coast line of  
55 1906 km. However, the possible abundance of marine actinobacterial resources remains  
56 largely unexplored, except some reports on the sponge-derived marine actinobacteria (Zhang  
57 *et al.*, 2006). During our screening of marine actinobacteria from the sediment samples of  
58 Dalian sea area, strain S187 was recovered from a sediment sample collected from Xinghai  
59 Bay. In this study, we performed polyphasic taxonomy on this strain, and proposed that strain  
60 S187<sup>T</sup> is a new species of the genus *Streptomyces*, for which the name *Streptomyces*  
61 *xinghaiensis* sp. nov. is proposed.

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63 Strain S187<sup>T</sup> was isolated from a marine sediment sample from Xinghai Bay, Dalian, China,  
64 after 2 weeks incubation at 28°C on Bennett's agar (Margalith & Beretta, 1960).  
65 Morphological, physiological and biochemical characterization of strain S187<sup>T</sup> were carried  
66 out following the standard protocol of the International *Streptomyces* Project (Shirling and  
67 Gottlieb, 1966; 1968 a, b, 1969). Colour determination was performed by comparing with the  
68 colour chips from ISCC-NBS COLOR CHARTS standard sample No. 2106 (Kelly, 1964).  
69 Spores and mycelia formation of strain S187<sup>T</sup> grown on ISP medium 2 at 28°C for 4 weeks  
70 were observed under light microscopy (Olympus microscope BH-2) and scanning electron  
71 microscopy (JSM-5600LV; JEOL). Phenotypic properties of strain S187<sup>T</sup> and its closely

72 related strains were examined using standard procedures (Shirling and Gottlieb, 1966;  
73 Williams *et al.*, 1983). The tolerance to NaCl was determined on ISP medium 2 after  
74 incubation at 28 °C for 3-4 weeks, whereas cell growth at the temperature ranging from 10 °C  
75 to 45 °C was examined for temperature tolerance. Antimicrobial activities of strain S187<sup>T</sup> and  
76 its closely related strains were tested as follows: Culture broth of the strains after cultivation in  
77 TSB medium at 28 °C for 6 days was centrifuged at 4 °C, 12000 rpm for 10 min, and 200 µl  
78 of the supernatant was applied to the agar blocks on the assay plates spread with test strains.  
79 Test strains were purchased from China General Microbiological Culture Collection Centre  
80 (CGMCC). The test strains are as following: *Escherichia coli* (CGMCC 1.797),  
81 *Staphylococcus aurea* (CGMCC 1.89), *Bacillus subtilis* (CGMCC 1.1849); *Pseudomonas*  
82 *aeruginosa* (CGMCC 1.2031), *Candida albicans* (CGMCC 2.538) and *Aspergillus niger*  
83 (CGMCC 3.2915).

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85 Amino acids and peptides in cell-wall hydrolysates were analysed by the methods described  
86 by Hasegawa *et al.* (1983). Whole cell sugars were analyzed by the methods of Lechevalier  
87 and Lechevalier (1980). Menaquinones and phospholipids were extracted from freeze dried  
88 cells and analyzed according to Collins (1985) and Lechevalier *et al.* (1977). Fatty acid  
89 composition was determined according to the procedure described elsewhere (Sasser, 1990).

90 Genomic DNA was extracted as described by Lee *et al.* (2003) and 16S rRNA gene was  
91 amplified according to Zhang *et al.* (2006). The DNA G+C content of the strain S187<sup>T</sup> was  
92 determined by using the HPLC method (Mesbah *et al.*, 1989). Closely related reference strains  
93 were chosen from BLAST (Altschul *et al.*, 1997) search results. Phylogenetic analysis was  
94 performed using the software package MEGA 2.1 version (Kumar *et al.*, 2001) after multiple

95 alignment by CLUSTAL X (Thompson *et al.*, 1997). A phylogenetic tree was reconstructed  
96 using neighbor-joining method (Saitou and Nei, 1987). The topology of the phylogenetic tree  
97 was evaluated by using the bootstrap resampling method of Felsenstein (1985) with 1000  
98 replicates.

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100 Morphological observation of a 28-day culture of strain S187<sup>T</sup> grown on yeast extract/malt  
101 extract agar (ISP medium 2) revealed that strain S187<sup>T</sup> had the typical characteristics of the  
102 genus *Streptomyces*. Aerial mycelium and substrate mycelium were well-developed without  
103 fragmentation. Long spore chains appeared as straight or flexuous, spores were smooth and  
104 non motile (Supplementary data Fig. S1). Strain S187<sup>T</sup> grew well between pH 6.0-9.0, with an  
105 optimum pH of 7.0-8.0. The range of temperature is 10-45 °C, with the optimum growth  
106 temperature at 28°C. Strain S187<sup>T</sup> grew in presences of 0-9% NaCl (w/v). Mycelia of strain  
107 S187<sup>T</sup> were well developed on some media tested, including yeast extract/malt extract agar  
108 (ISP medium 2), oatmeal agar (ISP medium 3), glycerol-asparagine agar (ISP medium 5),  
109 nutrient agar, and inorganic salt-starch agar (ISP medium 4), but the growth was poor on  
110 Czapek's agar and potato dextrose agar (Table 1). No diffusible pigments were observed on all  
111 test media.

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113 The cell wall of strain S187<sup>T</sup> contained LL-diaminopimelic acid and glycine, which indicates  
114 that this strain has a cell wall chemotype I (Lechevalier & Lechevalier, 1970). Whole-cell  
115 hydrolysates mainly contain ribose, glucose, galactose and small amount of mannose. The  
116 phospholipids of strain S187<sup>T</sup> were revealed to be phosphatidylethanolamine,

117 phosphatidylinositol, phosphatidylglycerol, phosphatidylinositol mannosides and one  
118 unknown phospholipid. The predominant menaquinones were MK-9 (H<sub>6</sub>)-40.8%, MK-9  
119 (H<sub>8</sub>)-38.2% and MK-9 (H<sub>2</sub>)-8.8%. The major fatty acids found were *iso-C*<sub>16:0</sub> (29.2%),  
120 *anteiso-C*<sub>15:0</sub> (14.0%), *antiso-C*<sub>17:0</sub> (11.6 %), *iso-C*<sub>14:0</sub> (8.8%), *iso-C*<sub>15:0</sub> (5.5%) and *iso-H C*<sub>16:1</sub>  
121 (4.6%), *anteiso-C C*<sub>17:1</sub> (4.0%), *C*<sub>16:0</sub> (4.0%) and *iso-C*<sub>17:0</sub> (3.5%). All the morphological  
122 characteristics and chemotaxonomic data showed that strain S187<sup>T</sup> should be assigned to the  
123 genus *Streptomyces*.

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125 The almost-complete 16S rRNA gene sequence (1490 nt) of strain S187<sup>T</sup> was aligned  
126 manually with corresponding 16S rRNA sequences of representative *Streptomyces* type strains  
127 retrieved from the GenBank/EMBL/DDBJ databases using BLAST search at  
128 <http://blast.ncbi.nlm.nih.gov/Blast.cgi> (Altschul *et al.*, 1997). Phylogenetic analyses based on  
129 16S rRNA gene sequence showed that the novel isolate fell into one distinct subclade with  
130 type strains of *Streptomyces flavofuscus* NRRL B-8036<sup>T</sup> (=DSM 41426<sup>T</sup>) and *Streptomyces*  
131 *albiaxialis* DSM 41799<sup>T</sup> with which it shares the 16S rRNA gene sequence similarity of  
132 98.1% and 97.5%, respectively. Another close strain is *Streptomyces ferralitis* SFOp68<sup>T</sup>, the  
133 16S rRNA gene sequence of which showed 95.6% similarity (1425 out of 1490 nucleotides)  
134 with that of S187<sup>T</sup>. The phylogenetic trees based on the 16S rRNA gene sequences of strain  
135 S187<sup>T</sup> and most closely related type strains of the genus *Streptomyces* are shown by  
136 neighbour-joining method (Fig. 1) and maximum parsimony method (Supplementary figure  
137 Fig. S2), respectively. DNA-DNA relatedness tests were performed between strain S187<sup>T</sup> and  
138 its closest match, *S. flavofuscus* DSM 41426<sup>T</sup> (=NRRL B-8036<sup>T</sup>) and *S. albiaxialis* DSM  
139 41799<sup>T</sup> using the optical renaturation method (De Ley *et al.*, 1970; Huss *et al.*, 1983; Jahnke

140 1992). The determined DNA-DNA relatedness values **between** S187<sup>T</sup> **and** strains *S.*  
141 *flavofuscus* NRRL B-8036 (DSM 41426<sup>T</sup>) and *S. albiacialis* DSM 41799<sup>T</sup> were 31.4% and  
142 46.9% respectively, which was significantly lower than 70%, the threshold value for the  
143 delineation of genomic species (Wayne *et al.*, 1987). Antimicrobial activities of the three  
144 strains were investigated, and it was showed that strain S187<sup>T</sup> displayed excellent activities  
145 against the four bacterial strains tested, while *S. flavofuscus* DSM 41426<sup>T</sup> **showed no activity**  
146 **against *P. aeruginosa* (Supplementary data Table S1)**. The phenotypic comparison between strain  
147 S187<sup>T</sup> and **its** closely related species of the genus *Streptomyces* were performed to  
148 differentiate **them** (Table 2). On the basis of morphological, chemotaxonomic and  
149 phylogenetic evidence and its physiological and **its** biochemical distinctiveness, it is  
150 confirmed that strain S187<sup>T</sup> should be a new member of the genus *Streptomyces*, for which the  
151 name *Streptomyces xinghaiensis* sp.nov. is proposed.

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### 153 **Description of *Streptomyces xinghaiensis* sp. nov.**

154 *Streptomyces xinghaiensis* (xing.hai.en'sis. **N.L. masc. adj.** xinghaiensis pertaining to Xinghai  
155 Bay of Dalian, China, the site from which the type strain was isolated).

156 Gram-positive, aerobic and non-motile. **Grows** well on glycerol aspaginate agar, Oatmeal agar,  
157 yeast extract/malt extract agar and inorganic salt-starch agar media, but poor on nutrient agar,  
158 potato dextrose agar and Czapek's agar media (Table 1). **Colonies** are whitish to **yellow**, ruffle.  
159 Aerial mycelium and substrate mycelium are well-developed without fragmentation. **Long**  
160 **spore chains appear as straight or flexuous, and spores are smooth with non-motile**. Casein,  
161 pectin and cellulose are degraded, but starch is not. Positive for catalase production and  
162 gelatin liquefaction. No soluble pigments are produced **on** all test media. Temperature range

163 for growth is 10 to 45°C with an optimum at 28°C. Growth occurs at pH 6.0-9.0 and in the  
164 presence of 0-9% NaCl (w/v). Good growth is found on almost all nitrogen sources tested, and  
165 can use D-glucose, sucrose, mannitol, fructose, mannose, galactose, ribose and rhamnose as  
166 carbon source, while cannot use trehalose, L-arabinose, inositol, raffinose, callobiose and  
167 sorbitol. More detailed phenotypic properties are mentioned in Table 2. Cell wall contains  
168 LL-diaminopimelic acid. Whole-cell hydrolysates mainly contain ribose, glucose, galactose  
169 and small amount of mannose. The chemotaxonomic properties are typical of the genus  
170 *Streptomyces*. The DNA G+C content of the type strain is 72.01 mol %.

171 The type strain, S187<sup>T</sup>(=NRRL B24674<sup>T</sup>= CCTCC AA 208049<sup>T</sup>= KCTC 19546<sup>T</sup>), was  
172 isolated from the marine sediment sample collected from Xinghai Bay, Dalian, Liaoning  
173 Province, China.

174

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**Table 1. Culture characteristics of strain S187<sup>T</sup> and its most closely related neighbours**

Culture medium	<i>S. singhaiensis</i> S187 <sup>T</sup>			<i>S. flavofuscus</i> DSM 41426 <sup>T</sup>			<i>S. albivialis</i> DSM 41799 <sup>T</sup>		
	Growth	Aerial mycelium	Substrate mycelium	Growth	Aerial mycelium	Substrate mycelium	Growth	Aerial mycelium	Substrate mycelium
Yeast extract/malt extract	Good	White	Dark orange yellow	Good	White	Dark orange yellow	Good	Strong olive green	Moderate reddish brown
Oatmeal agar	Good	White	Yellow white	Good	White	Strong olive green	Good	White	Deep orange yellow
Potato Dextrose agar	Poor	-	Pale orange yellow	Good	White	Yellow white	Good	Vivid bluish green	Moderate reddish brown
Nutrient agar	Moderate	White	Light orange yellow	Good	White	Moderate orange yellow	Poor	-	Light orange yellow
Czapek's agar	Poor	-	Light yellow brown	moderate	White	Strong olive green	Good	Strong olive green	Light yellow brown
Glucose/Asparagine agar	Good	Vivid bluish green	Yellow white	Good	White	Pale orange yellow	Good	Vivid yellowish pink	Moderate reddish brown
Inorganic salts starch agar	Good	White	Pale orange yellow	Good	White	Yellow white	Good	Deep olive green	Pale orange yellow

Note: '-', no growth. Diffusible pigment was not observed on any of the media listed. Colors were taken from ISCC-NBS COLOR CHARTS standard sample No. 2106 (Kelly, 1964). Plates were incubated for 4 weeks at 28°C.

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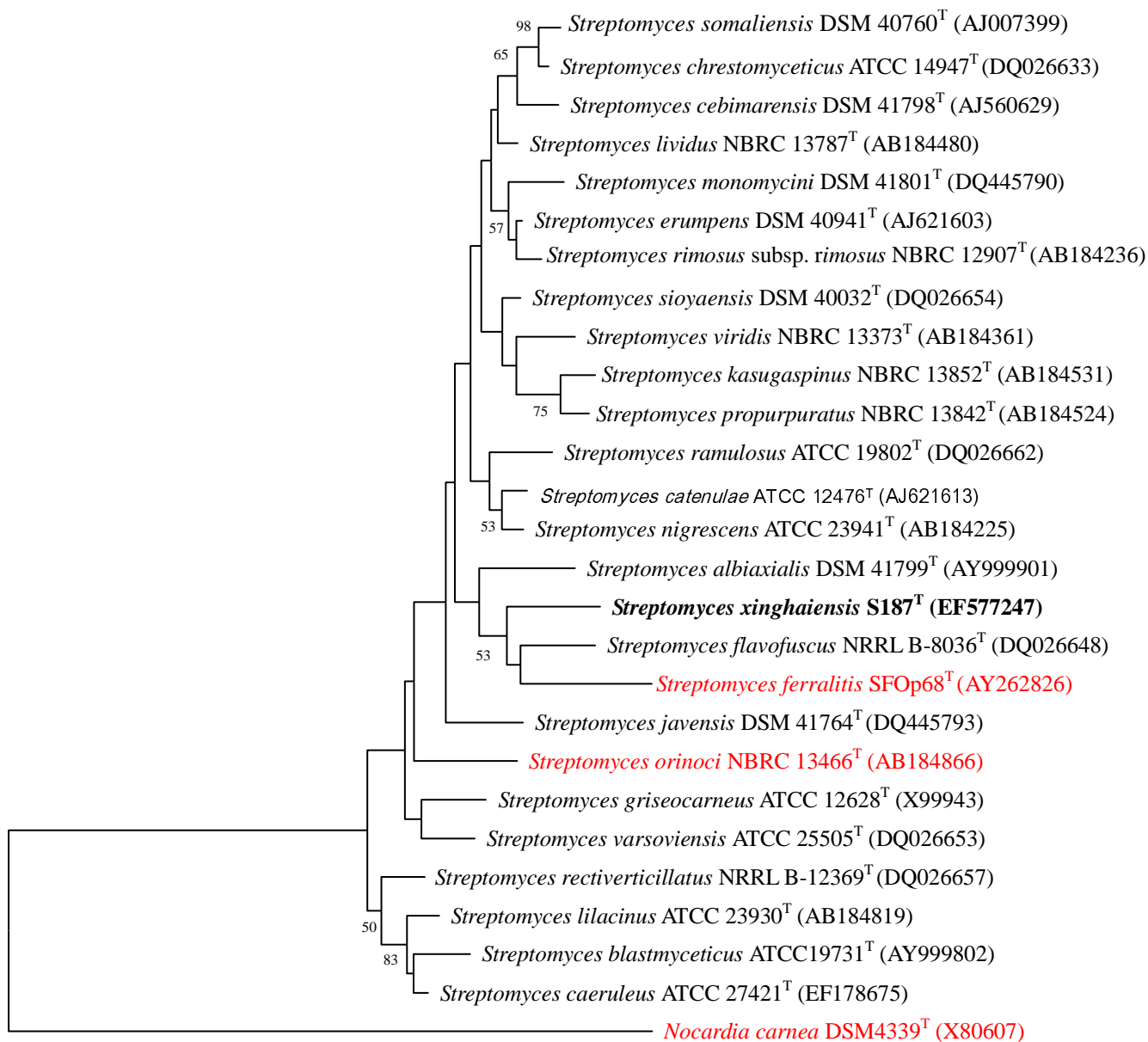
**Table 2. Physiological and biochemical features that differentiate strain S187<sup>T</sup> from closely related *Streptomyces* species**

1, Strain S187<sup>T</sup>; 2, *S. flavofuscus* DSM 41426<sup>T</sup>; 3, *S. Albiaxialis* DSM 41799<sup>T</sup>.  
+; Positive, -; Negative. **Only the items that showed differences in the strains are presented.**

<b>Characteristics</b>	<b>1</b>	<b>2</b>	<b>3</b>
<b>Degradation</b>			
Casein	+	+	-
Cellulose	+	-	+
<b>Tolerance to NaCl (% w/v)</b>			
	0-9	0-10	0-6
<b>Optimum NaCl (% v/v)</b>			
	0-5	0-5	0-3
<b>Growth at 5°C</b>			
	-	-	+
<b>Growth at pH</b>			
	7-9	6-9	7-9
<b>Carbon utilization</b>			
L-arabinose	-	-	+
ribose	+	+	-
<i>i</i> -inositol	-	-	+
L-rhamnose	+	+	-
callobiose	-	+	+
sorbitol	-	-	+
<b>Nitrogen utilization</b>			
lysine	-	+	-
phenylalanine	+	-	+
serine	+	+	-
xanthine	+	+	-

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282 **Fig. 1. Phylogenetic tree based on 16S rRNA gene sequence analysis and constructed**  
 283 **using the neighbor-joining method showing the position of strain S187<sup>T</sup> among its**  
 284 **phylogenetic neighbors. Numbers at the branch nodes are bootstrap values, expressed as**  
 285 **a percentage of 1000 replicates (only values above 50% are shown). Bar 0.01**  
 286 **substitutions per nucleotide position.**



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## Supplementary data

**Table S1. Antimicrobial activities of S187<sup>T</sup> and closely related strains *S. flavofuscus* DSM 41426<sup>T</sup> and *S. albiaxis* DSM 41799<sup>T</sup>. The value indicated the inhibition zone diameter (mm) against the test strain.**

Test strains	S187 <sup>T</sup>	<i>S. flavofuscus</i> DSM 41426 <sup>T</sup>	<i>S. albiaxis</i> DSM 41799 <sup>T</sup>
<i>E. coli</i>	31	19	-
<i>S. aureus</i>	30	19	-
<i>B. subtilis</i>	38	21	-
<i>P. aeruginosa</i>	22	-	-
<i>C. albicans</i>	-	-	-
<i>A. niger</i>	-	-	-

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**Fig. S1. Scanning electron micrograph indicates the spore chain morphology of strain S 187<sup>T</sup> grown on ISP medium 2 for 4 weeks at 28 °C. Bar 5 μm.**

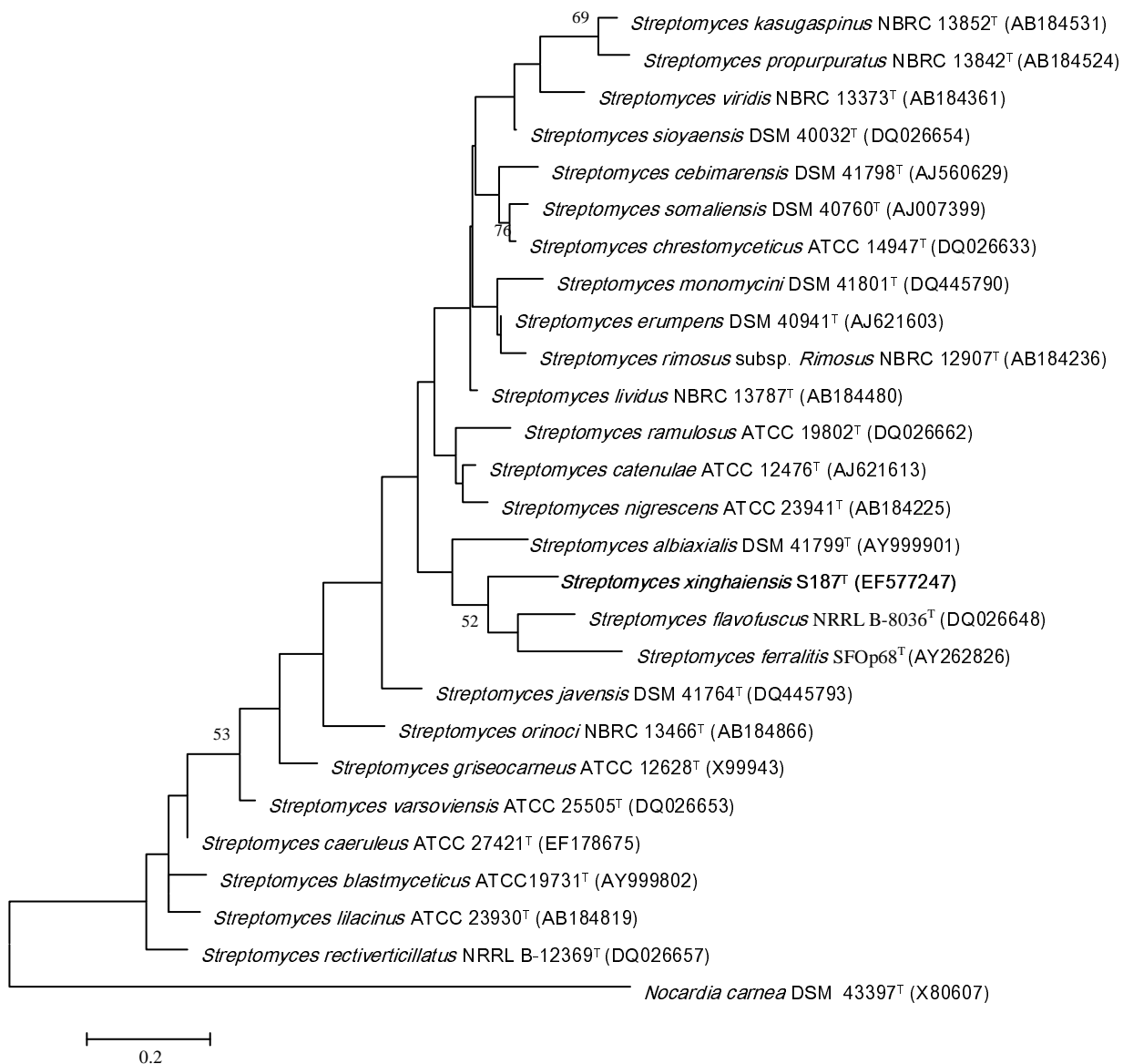


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**Fig. S2. Phylogenetic tree based on 16S rRNA gene sequence analysis and constructed using the maximum parsimony method showing the position of strain S187<sup>T</sup> among its phylogenetic neighbors. Numbers at the branch nodes are bootstrap values, expressed as a percentage of 1000 replicates (only values above 50% are shown). Bar 0.02 substitutions per nucleotide position.**



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