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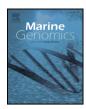
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#### 1 Genomics/technical resources

# Draft genomic DNA sequence of strain *Halomonas* sp. FS-N4 exhibiting high catalase activity

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#### ABSTRACT

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#### 28 1. Introduction

Since genus Halomonas had been organized as a genus in 1980 29(Vreeland et al., 1980), there was no report about any members 30whose catalase activity was above 1 katal/mg. Recently, we isolated a 31 strain, FS-N4, which can grow in the medium Marine Broth 2216 32 (Difco; MB) with initial hydrogen peroxide concentration of 5 M, 33 shows a strong oxidation resistance, and the cell-free extract enzyme 34 catalase activity can reach 13.33 katal/mg. To attain deeper insight, 35 a whole-genome sequence of the strain FS-N4 was established, the 36 genome sequence may provide a basis to improve the growth of the 37 38 strain FS-N4, and the probable industrial application.

The seawater was collected in the China East Sea. near the Zhoushan 39 City, China. The seawater (5 mL) was added to 50 mL MB, and incubated 40at 28 °C for 2 days. The cultures were added to the fresh medium with 41 425 mM hydrogen peroxide by 10% volume. After cultured at 28 °C for 2 days, the cultures were added to the fresh medium with 10 mM 43 hydrogen peroxide. By repeated selection with increasing amounts of 44 45 hydrogen peroxide, the initial concentration reached 5 M. After spreading on plate without hydrogen peroxide for purifying, strain FS-N4 46 (=CGMCC 9352) was isolated. 47

The cells were collected and disrupted in the phosphate buffer (same volume of the culture broth) by ultrasonic wave, cell-free extracts were harvested by centrifugation. Catalase activity was measured spectrophotometrically by monitoring the decrease in absorbance at 240 nm caused by the disappearance of hydrogen peroxide (Beers and Sizer, 1952),

http://dx.doi.org/10.1016/j.margen.2014.08.002 1874-7787/© 2014 Elsevier B.V. All rights reserved. Halomonas sp. FS-N4 is a bacterium, which can grow in the medium Marine Broth 2216 with 5 M initial hydrogen17peroxide concentration, shows a strong oxidation resistance, and the crude enzyme activity can reach as high as1813.33 katal/mg. We reported the draft genome sequence of H. sp. FS-N4, showing that it contains 3434 protein-19coding genes, including the genes putatively involved in the response to the oxidative stress, among which a20phytochrome-like gene might be a key point to survive in the environment with high concentration of hydrogen21peroxide and exhibit high catalase activity.22

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using a spectrophotometer (DU 800; BECKMAN). The  $\varepsilon$  at 240 nm for 53 hydrogen peroxide was assumed to be 43.6 M<sup>-1</sup>·cm<sup>-1</sup> (Hildebrandt 54 and Roots, 1975). After cultured for 27 h, catalase activity of the strain 55 FS-N4 reached the peak, 13.33 katal/mg (=79997.36 U/mg; the amount 56 of enzyme that decomposed 1 µmol of hydrogen peroxide per minute 57 was defined as 1 U of activity). Catalase activity in the cell-free extracts 58 of the strain FS-N4 and other typical catalase producers were showed 59 in Table 1. The specific activity of the catalase of the strain FS-N4 was 60 more than 2.5-fold that of the catalase of *Rhizobium radiobacter* 2-1, 61 which exhibits the highest activity shown in the references (Nakayama 62 et al., 2008). 63

Genomic DNA sequencing of strain FS-N4 was performed using 64 Solexa paired-end sequencing technology (HiSeq 2000 System, Illumina, 65 Inc., USA) (Bentley et al., 2008) with a whole-genome shotgun (WGS) 66 strategy, with a 500 bp-span paired-end library (546 Mb available 67

Table 1	t1.1
Catalase activities in cell extracts of <i>Halomonas</i> sp. FS-N4 and other catalase producers.	t1.2

Strain or species	Activity $(U \cdot mL^{-1})$	Protein $(mg \cdot mL^{-1})$	Specific activity $(U \cdot mg^{-1})$	t1.3
H. sp. FS-N4	75,523	0.9	79,997	t1.4
Strain 2-1ª	87,594	2.9	31,503	t1.5
M. luteus <sup>a</sup>	36,986	3.4	11,365	t1.6
P. fluorescens <sup>a</sup>	892	6.8	133	t1.7
R. radiobacter (A. tumefaciens) <sup>a</sup>	129	2.6	50	t1.8
E. coli <sup>a</sup>	117	3.5	36	t1.9

<sup>a</sup> Data were taken from Nakayama et al., 2008. Cells were suspended in the buffer of 1/5 t1.10 amount of the culture broth. t1.11

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#### 2

t2.1	Table 2	
	-	

t2.2 Genome features.

t2.3		Halomonas sp. FS-N4
t2.4	Size (bp)	3,797,897
t2.5	Contigs	20
t2.6	GC content (%)	51.87
t2.7	Coding (%)	88.93
t2.8	CDS	3434
t2.9	rRNA genes	6
t2.10	tRNA genes	60
t2.11	16S rDNA identity <sup>a</sup>	98.8

t2.12  $^{a}$  Compared to the type strain *Halomonas andesensis* LC6<sup>T</sup> (Daniel et al., 2010).

reads). All these clean reads were assembled into 20 scaffolds with total 68 3,797,897 bp (coverage: 142.9×) using the Velvet 1.2.07 (Zerbino et al., 69 2009). The detail of FS-N4 genomic sequencing results was showed in 70 Table 2. The results were extracted using Rapid Annotation using Sub-71 system Technology (RAST) (Aziz et al., 2008), and functions of the gene 72 products were annotated by the same program. 73

This draft genome shotgun project has been deposited as a primary 74 project at DDBJ BioProject (the accession number: PRJNA241396). The 75 draft genome sequence of the strain FS-N4 was deposited in the 76 GenBank database under the accession number JHQL00000000. 77

The GenBank accession number for the 16S rRNA gene sequence of 78 strain FS-N4 is KM079655. Neighbor-joining phylogenetic tree based 79 on the 16S rRNA gene of FS-N4 and related species was showed in 80 Fig. 1. According to the tree, strain FS-N4 shared the highest sequence 81

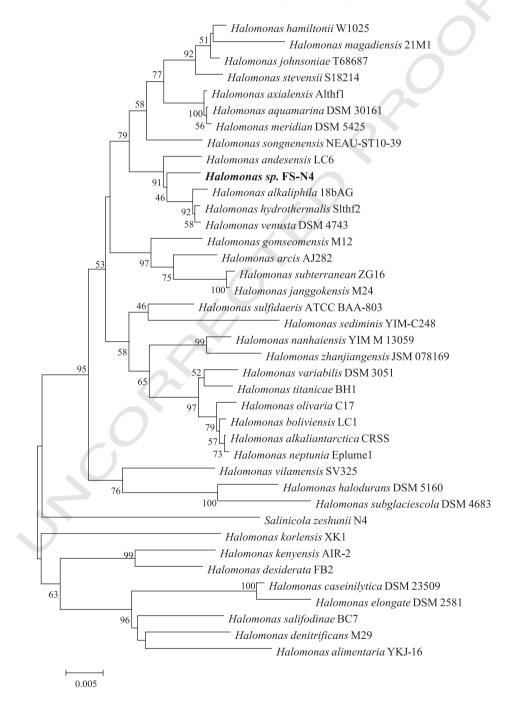


Fig. 1. Neighbor-joining phylogenetic tree based on 16S rRNA gene sequences showing the relationship of strain FS-N4 and other species of the genus *Halomonas*. Numbers at branch points are bootstrap values (percentages based on 1000 replications; only showed values above 45% are shown). Bar, 0.5% substitution per nucleotide position.

## **Q2** similarity of 98.8% with *Halomonas andesensis* LC6<sup>T</sup>, but did not cluster with it in the phylogenetic tree. It showed ambiguous taxonomic status of strain FS-N4, so we named it *H*. sp. FS-N4.

03 Bioinformatics analyses use Basic Local Alignment Search Tool (BLAST) (Altschul et al., 1997) and RAST. The analyzed results were 86 showed in Fig. S1 and also could be found on the web (http://rast. 87 nmpdr.org/rast.cgi?page=JobDetails&job=140167), demonstrated 88 that the H. sp. FS-N4 genome contained genes coding for 24 oxidative 89 90 stress related proteins. Among them, the only superoxide dismutase 91shared the highest similarity with manganese superoxide dismutase 92(sodA), which was the enzyme that catalyzes the dismutation of superoxide  $(O_2^-)$  into oxygen and hydrogen peroxide (Keele et al., 93 941970), and two of catalases were most similar to hydroperoxidase 95I (catalase, katG), Manganese containing catalase, which was an important antioxidant enzyme that catalyzes decomposition and 96 disproportionation of hydrogen peroxide, respectively (Chelikani et al., 97 2004), forming dioxygen and water. The other catalase was most similar 98 to hydroperoxidase II (catalase, *katE*), which were haem-containing 99 enzymes that use hydrogen peroxide as the electron acceptor to 100 catalyze a number of oxidative reactions (Nelson et al., 1994). 101

Moreover, FS-N4 genome contained genes coding for proteins regulating the responses to hydrogen peroxide  $(H_2O_2)$  and superoxide  $(O_2^-)$ , including alkyl hydroperoxide reductase subunits (*ahpC* and *ahpF*), glutaredoxin I (*grxA*), glutathione reductase (*gorA*), Fur repressor, Zinc uptake regulation protein ZUR, and peptide methionine sulfoxide reductase (*gsrA*).

Alkyl hydroperoxide reductase (Ahp), KatG and KatE were the most 108 109important proteins in the process of scavenging hydrogen peroxide in vivo (Seaver and Imlay, 2001). The thiol-based peroxidase Ahp 110 consisted of two subunits, AhpC and AhpF, it transfered electrons from 111 NADH to H<sub>2</sub>O<sub>2</sub> and reduced H<sub>2</sub>O<sub>2</sub> to water. Fur repressor and Zinc 112 113uptake regulation protein ZUR were both involved in the PerR regulon, which was known to be highly induced by oxidative stress caused by 114 hydrogen peroxide or paraquat. 115

According to the annotation results of RAST, the genes related to the 116 oxidative-stress-inducible activities were compared with those of 117 Hestiasula zhejiangensis, the results showed that the related genes were 04 119 almost the same, except a phytochrome-like gene. As the definite enhancement by phytochrome of the catalase level was demonstrated 120in mustard (Drumm and Schopfer, 1974) and the induction of the Cat3 121 expression is probably regulated by a very low fluence phytochrome 122123 response (Polidoros and Scandalios, 1997), the phytochrome-like gene might be an important gene for strain FS-N4 to survive in the high-124 hydrogen-peroxide environment and produce high-catalase-activity 125extract. It needed more works to reveal it. 126

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### Supplementary data to this article can be found online at http://dx. 127

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